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## **Pharmacogenomics Impact on Drug Development**

*Looking Through a Window  
of the FDA*

*SARS Virus Inhibited by siRNA*

# Multicolor In Vivo Imaging in Mouse Models of Cancer

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## FLUORESCENT PROTEINS

Cloning of the reporter gene green fluorescent proteins (GFP) from the jellyfish *Aequorea victoria* in 1992 began the era of visualization of cellular and subcellular processes in live cells in real time (1). Most importantly, it was shown that GFP could be highly expressed as a transgene in many types of organisms (2,3). Numerous cellular processes that could previously only be studied biochemically outside the cell or in fixed cells could now be visualized in real time in living cells. This powerful reporter gene has been genetically modified to increase its fluorescence intensity and to fluoresce at other distinct wavelengths, such as blue and yellow (4–7), and humanized for high expression and strong signal (8). GFP requires no other *Aequorea* proteins, substrates, or cofactors to fluoresce (9). Recently, other fluorescent proteins have been isolated from various organisms, in particular the coral *Discosoma*, which is the source of the brilliant red fluorescent protein (RFP) DsRed, that has been genetically modified to DsRed-2 (10). The availability of multicolored spectrally distinct fluorescent proteins enables simultaneous visualization of multiple processes in living cells.

## IN VIVO IMAGING

Our laboratory pioneered the use of fluorescent proteins for in vivo imaging, where their very high levels of fluorescence have enabled whole-body imaging of primary and metastatic tumor growth (11), angiogenesis (12), gene expression (13), stem cell and other cell tracking (14), and infection in intact animals (15). In order to externally image and follow the natural course or inhibition of tumor progression and metastasis or other processes outlined above in vivo, high specificity, a strong signal, high resolution, and good physiological conditions are necessary. In vivo imaging with fluorescent proteins is possible without

anesthetizing the animal or the injection of any type of substrate. Images can be captured from fluorescent proteins in numerous organs of the body in freely moving animals. The technology is based on the bright intrinsic fluorescence of GFP and RFP, which is partly caused by the high quantum yield of these fluorophores (7,10). The use of narrow band-pass or spectrally resolving filters can fully eliminate any autofluorescence background in the animal.

## TECHNIQUES FOR IN VIVO IMAGING

External whole-body imaging of mice with primary and metastatic tumors, which are genetically labeled with the fluorescent proteins GFP and RFP, is a simple but powerful tool for investigating tumor development. For tumor cells to be visualized with this technique, they must be transduced with GFP or RFP genes, such that they become brightly fluorescent. This can be accomplished by in vitro (11,16) and in vivo (17) selection of such fluorescent tumor cells.

To produce metastasis in mice, the genetically fluorescent tumors should be transplanted orthotopically (12,16,18–26). Once the GFP- or RFP-expressing tumors have developed and metastases have formed, they can be visualized in the live mouse by use of whole-body imaging with fairly simple equipment.

A fluorescence light box, with fiber-optic lighting at about 470 nm and appropriate filters placed on top of the light box, can be used to image large tumors and can be viewed with the naked eye (11). Alternatively, the light box can be linked to a charge-coupled device (CCD) camera with appropriate filters to enable images to be captured digitally by a computer, displayed on a monitor, and digitally stored (11).

In order to visualize smaller tumors and metastases, the animal can be put on a fluorescence dissecting microscope, which incorporates a light

lows unrestrained animals to be imaged without any perturbation or substrate—irradiation with nondamaging blue light is the only step needed. Images can be captured with a fairly simple apparatus, and there is no need for absolute darkness. The magnitude of bioluminescence measured in vivo varies with time after the injection and dosage of luciferin, which makes repeatable quantification very different (36).

The multicolored spectrally distinct set of fluorescent proteins is rapidly growing, especially those that emit at longer wavelengths. The multicolor fluorescent proteins form the basis of the most versatile, simple, highest-resolution, and most sensitive in vivo imaging technology. It is expected that new fluorescent proteins emitting close to 700 nm will soon be on the horizon and suitable for very deep in vivo multicolor imaging.

The portfolio of multicolor fluorescent proteins enables a second new era of biological visualization, that of in vivo imaging at the cellular level. With fluorescent proteins, multiple cellular processes can be simultaneously visualized in the intact animal that previously could only be seen in cultured cells or detected biochemically outside of the cell. The application of this imaging technology for drug discovery and development and the understanding of disease is possible by linking specific processes to expression of specific colored fluorescent proteins in any type of animal model for external noninvasive imaging.

REFERENCES

- Prasher, D.C., V.K. Eckenrode, W.W. Ward, F.G. Prendergast, and M.J. Cormier. 1992. Primary structure of the *Aequorea victoria* green-fluorescent protein. *Gene* 111:229-233.
- Chalfie, M., Y. Tu, G. Euskirchen, W.W. Ward, and D.C. Prasher. 1994. Green fluorescent protein as a marker for gene expression. *Science* 263:802-805.
- Cheng, L., J. Fu, A. Tsukamoto, and R.G. Hawley. 1996. Use of green fluorescent protein variants to monitor gene transfer and expression in mammalian cells. *Nat. Biotechnol.* 14: 606-609.
- Cormack, B., R. Valdivia, and S. Falkow. 1996. FACS-optimized mutants of the green fluorescent protein (GFP). *Gene* 173:33-38.
- Cramer, A., E.A. Whitehorn, E. Tate, and W.P.C. Stemmer. 1996. Improved green fluorescent protein by molecular evolution using DNA shuffling. *Nat. Biotechnol.* 14:315-319.
- Delagrave, S., R.E. Hawtin, C.M. Silva, M.M. Yang, and D.C. Youvan. 1995. Red-shifted excitation mutants of the green fluorescent protein. *Biotechnology* 13:151-154.
- Heim, R., A.B. Cubitt, and R.Y. Tsien. 1995. Improved green fluorescence. *Nature* 373:663-664.
- Zolotukhin, S., M. Potter, W.W. Hauswirth, J. Guy, and N. Muzyczka. 1996. A "humanized" green fluorescent protein cDNA adapted for high-level expression in mammalian cells. *J. Virol.* 70:4646-454.
- Morin, J. and J. Hastings. 1971. Energy transfer in a bioluminescent system. *J. Cell Physiol.* 77:313-318.
- Matz, M.V., A.F. Fradkov, Y.A. Labas, A.P. Savitsky, A.G. Zaraisky, M.L. Markelov, and S.A. Lukyanov. 1999. Fluorescent proteins from nonbioluminescent *Anthozoa* species. *Nat. Biotechnol.* 17:969-973.
- Yang, M., E. Baranov, P. Jiang, F.X. Sun, X.M. Li, L. Li, et al. 2000. Whole-body optical imaging of green fluorescent protein-expressing tumors and metastases. *Proc. Natl. Acad. Sci. USA* 97:1206-1211.
- Yang, M., E. Baranov, X.M. Li, J.W. Wang, P. Jiang, L. Li, et al. 2001. Whole-body and intravital optical imaging of angiogenesis in orthotopically implanted tumors. *Proc. Natl. Acad. Sci. USA* 98:2616-2621.
- Yang, M., E. Baranov, A.R. Moossa, S. Penman, and R.M. Hoffman. 2000. Visualizing gene expression by whole-body fluorescence imaging. *Proc. Natl. Acad. Sci. USA* 97:12278-12282.
- Li, L., J. Mignone, M. Yang, M. Matic, S. Penman, G. Enkolopov, and R.M. Hoffman. 2003. Nestin expression in hair follicle sheath progenitor cells. *Proc. Natl. Acad. Sci. USA* 100: 9958-9961.
- Zhao, M., M. Yang, E. Baranov, X. Wang, S. Penman, A.R. Moossa, and R.M. Hoffman. 2001. Spatial-temporal imaging of bacterial infection and antibiotic response in intact animals. *Proc. Natl. Acad. Sci. USA* 98:9814-9818.
- Chishima, T., Y. Miyagi, X. Wang, H. Yamaoka, H. Shimada, A.R. Moossa, and R.M. Hoffman. 1997. Cancer invasion and micrometastasis visualized in live tissue by green fluorescent protein expression. *Cancer Res.* 57:2042-2047.
- Hasegawa, S., M. Yang, T. Chishima, Y. Miyagi, H. Shimada, A.R. Moossa, and R.M. Hoffman. 2000. In vivo tumor delivery of the green fluorescent protein gene to report future occurrence of metastasis. *Cancer Gene Ther.* 7:1336-1340.
- Chishima, T., Y. Miyagi, X. Wang, E. Baranov, Y. Tan, H. Shimada, et al. 1997. Metastatic patterns of lung cancer visualized live and in process by green fluorescent protein expression. *Clin. Exp. Metastasis* 15:547-552.
- Yang, M., S. Hasegawa, P. Jiang, X. Wang, Y. Tan, T. Chishima, et al. 1998. Widespread skeletal metastatic potential of human lung cancer revealed by green fluorescent protein expression. *Cancer Res.* 58:4217-4221.
- Rashidi, B., M. Yang, P. Jiang, E. Baranov, Z. An, X. Wang, et al. 2000. A highly metastatic Lewis lung carcinoma orthotopic green fluorescent protein model. *Clin. Exp. Metastasis* 18:57-60.
- Hastings, R.H., D.W. Burton, R.A. Quintana, E. Biederman, A. Gujral, and L.J. Deftos. 2001. Parathyroid hormone-related protein regulates the growth of orthotopic human lung tumors in athymic mice. *Cancer* 92:1402-1410.
- Yang, M., P. Jiang, F.X. Sun, S. Hasegawa, E. Baranov, T. Chishima, et al. 1999. A fluorescent orthotopic bone metastasis model of human prostate cancer. *Cancer Res.* 59:781-786.
- Glinkii, A.B., B.A. Smith, P. Jiang, X.-M. Li, M. Yang, R.M. Hoffman, and G.V. Glinksky. 2003. Viable circulating metastatic cells produced in orthotopic but not ectopic prostate cancer models. *Cancer Res.* 63:4239-4243.
- Bouvet, M., M. Yang, S. Nardin, X. Wang, P. Jiang, E. Baranov, et al. 2000. Chronologically-specific metastatic

- targeting of human pancreatic tumors in orthotopic models. *Clin. Exp. Metastasis* 18:213-218.
25. **Bouvet, M., J.W. Wang, S.R. Nardin, R. Nassirpour, M. Yang, E. Baranov, et al.** 2002. Real-time optical imaging of primary tumor growth and multiple metastatic events in a pancreatic cancer orthotopic model. *Cancer Res.* 62:1534-1540.
  26. **Yang, M., T. Chishima, X. Wang, E. Baranov, H. Shimada, A.R. Moossa, R.M. Hoffman.** 1999. Multi-organ metastatic capability of Chinese ovary cells revealed by green fluorescent protein (GFP) expression. *Clin. Exp. Metastasis* 17:417-422.
  27. **Yang, M., E. Baranov, J.W. Wang, P. Jiang, X. Wang, F.X. Sun, et al.** 2002. Direct external imaging of nascent cancer, tumor progression, angiogenesis, and metastasis on internal organs in the fluorescent orthotopic model. *Proc. Natl. Acad. Sci. USA* 99:3824-3829.
  28. **Yamamoto, N., M. Yang, P. Jiang, M. Xu, H. Tsuchiya, K. Tomita, et al.** 2003. Real-time imaging of individual color-coded metastatic colonies in vivo. *Clin. Exp. Metastasis* 20: 633-638.
  29. **Yamamoto, N., M. Yang, P. Jiang, M. Xu, H. Tsuchiya, K. Tomita, et al.** 2003. Determination of clonality of metastasis by cell-specific color-coded fluorescent-protein imaging. *Cancer Res.* 63:7785-7790.
  30. **Yang, M., L. Li, P. Jiang, A.R. Moossa, S. Penman, and R.M. Hoffman.** 2003. Dual-color fluorescence imaging distinguishes tumor cells from induced host angiogenic vessels and stromal cells. *Proc. Natl. Acad. Sci. USA* 100:14259-14262.
  31. **Cody, C.W., D.C. Prasher, W.M. Westler, F.G. Prendergast, and W.W. Ward.** 1993. Chemical structure of the hexapeptide chromophore of the *Aequorea* green fluorescent protein. *Biochemistry* 32:1212-1218.
  32. **Levenson, R., M. Yang, and R.M. Hoffman.** 2003. Whole-body spectral imaging of fluorescently labeled orthotopic lung tumors in the live mouse. *Proc. Am. Assoc. Cancer Res.* 44:776.
  33. **Ilyin, S.E., M.C. Flynn, and C.R. Plata-Salaman.** 2001. Fiber-optic monitoring coupled with confocal microscopy for imaging gene expression in vitro and in vivo. *J. Neurosci. Methods* 108:91-96.
  34. **Sweeney, T.J., V. Mailander, A.A. Tucker, A.B. Olomu, W. Zhang, Y. Cao, et al.** 1999. Visualizing the kinetics of tumor-cell clearance in living animals. *Proc. Natl. Acad. Sci. USA* 96: 12044-12049.
  35. **Contag, C.H., D. Jenkins, P.R. Contag, and R.S. Negrin.** 2000. Use of reporter genes for optical measurements of neoplastic disease in vivo. *Neoplasia* 2:41-52.
  36. **Burgos, J.S., M. Rosol, R.A. Moats, V. Khankaldyyan, D.B. Kohn, M.D. Nelson, Jr., and W.E. Laug.** 2003. Time course of bioluminescent signal in orthotopic and heterotopic brain tumors in nude mice. *BioTechniques* 34:1184-1188.

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