Efficacy of intra-hepatectomy 5-FU on recurrence and metastasis of human hepatocellular carcinoma in nude mice

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The aim of our study was to evaluate the efficacy of intra-operative 5-FU for the treatment of highly metastatic HCC in an orthotopic nude mouse model.

MATERIAL AND METHODS

Animals

Athymic nu/nu Balb/c mice of both sexes, 6–7 weeks old, were used in our study. The animals were maintained in a specific pathogen-free environment in compliance with USPHS guidelines governing the care and maintenance of experimental animals. All animal studies were conducted in accordance with the principles and procedures outlined in the National Institutes of Health Guide for the Care and Use of Animals under assurance number A3873-1. Mice were fed with an autoclaved laboratory rodent diet (Tecklad LM-485, Western Research Products, Orange, CA).

Human hepatocellular carcinoma

A highly metastatic human hepatocellular carcinoma nude mouse model, LCI-D20, was previously established by surgical orthotopic implantation (SOI).28

Tumor implantation on the nude mice liver

In the present study, LCI-D20 human hepatoma tissue fragments were implanted by SOI directly on the left lobe of the liver in 60 animals using the following methods: Liver metastases of LCI-D20 that originated from SOI to the liver of nude mice were harvested and carefully inspected under a dissecting microscope (5×) to remove necrotic tissue. The harvested tumor tissues were then equally divided into small pieces of 1 mm³ each. Tumor tissue fragments were mixed thoroughly before the implantation procedure to ensure that each mouse received equally viable tissues. The left lobe of the liver of the mice was isolated via a left subcostal incision under isoflurane anesthesia. A small cut was then made on the glissonian capsule. Three 1 mm³ pieces of the above tumor tissue fragments were inserted into the incision on the left lobe and then fixed in place using an 8-0 nylon suture. The abdomen was closed with a 6-0 silk suture using different instruments than those used for tumor implantation in order to prevent any tumor cell seeding at the incision site.

Partial hepatectomy

Mice were anesthetized with isoflurane inhalation and put in a supine position. The abdomen was sterilized with iodine and alcohol swabs. To prevent any residual tumor growth, the left subcostal incision site, which was used for tumor implantation, was completely excised. Through this abdominal wall opening, the tumor-bearing left lobe of the liver was isolated and its biliary-vascular bundle ligated with a 6-0 nylon suture. Then the entire left...
lobe was resected. The incision line was at least 1 cm distant from the liver metastasis. The resected lobe corresponded to approximately 30% of the total liver. The abdomen was then closed with a 6-0 silk suture using different instruments than those used for the resection to prevent any tumor spreading at the incision site.

Chemotherapeutic study design
5-FU (Calbiochem-Novabiochem, San Diego, CA) was used at 20 mg/kg/0.5 ml i.p. There were 6 groups of animals in this study. In group 1, the control animals were treated with saline, 0.5 ml, once a day for 5 days. In group 2, animals were treated with 5-FU only once a day for 5 days, with the first dose starting 3 days after tumor implantation. In group 3, a curative liver resection was performed 3 days after tumor implantation. In group 4, which received adjuvant treatment, animals were first given a curative liver resection, then after 2 days of rest, were subsequently treated with 5-FU once a day for 5 days. In group 5, which received neoadjuvant treatment, animals were treated with 5-FU once a day for 5 days, then after 2 days of rest, a curative liver resection was performed. In group 6, intra-hepatic chemotheraphy (IHC), the first 5-FU dose was given to the animals 2 hr prior to liver resection. Four more daily doses of 5-FU were given to the animals beginning from the first post-operative day (Table I).

Statistical analysis
The incidence of survival at defined time points and the incidence of metastasis was analyzed using the Fisher exact test. Median survival was analyzed using the Wilcoxon rank-sum test.

RESULTS

Survival
All animals were sacrificed when they became moribund. Five animals in the IHC treatment group did not show any sign of neoplastic disease and were sacrificed at day-150 post-tumor implantation. Animals with IHC survived much longer than all other treatment or control groups (Fig. 1). The median survival of the IHC group was 127 days compared with 53 days for the control group (p < 0.006). The median survival of animals in the neoadjuvant group was 78 days (p < 0.021 compared with the control group). Though survival of animals in the surgery-alone, 5-FU-alone and adjuvant post-operative 5-FU groups was longer than the control group, it was not statistically significant. When all animals with neoadjuvant 5-FU treatment had died, 60% of animals with IHC were still alive (p < 0.011) (Table II).

Systemic metastasis
All animals were explored for metastasis at necropsy. In the control group, 100% of the animals had metastasis in lymph nodes draining the liver. In animals treated with 5-FU alone, 100% of the animals had lymph node metastases. In animals treated with surgical resection only without chemotherapy, 75% of the animals had lymph node metastases. In animals treated with surgery and subsequent chemotherapy (IHC), 63% of the animals had lymph node metastases. In the neoadjuvant chemotherapy group, 63% of the animals had lymph node metastases. In the IHC group, only 2 animals had lymph node metastases (20%). Peritoneal carcinomatosis occurred in 67% of animals in the control group and in only 10% of animals treated with IHC (p < 0.020). Five mice treated with IHC sacrificed at day-150 post-tumor implantation were found free of neoplastic disease. All other animals in all groups had recurrences found in the liver. The metastatic patterns and frequency in the different treatment groups are shown in Table III.

DISCUSSION
Adjuvant, post-operative and standard chemotherapeutic regimes have not significantly improved survival of patients with HCC. Early recurrence in the remnant liver after a curative liver resection is high. The reasons for recurrences in the residual liver after a curative liver resection are believed to be due to microscopic residual foci of tumor or spreading of malignant cells by surgical manipulation or both. It is known that surgical manipulation spreads HCC cells into the portal circulation. Pharmacokinetic and phase-1 clinical studies have indicated that i.p. administration of 5-FU achieves drug concentrations in the peritoneal cavity between 300 and 2,200 times higher than in the systemic circulation. Anatomic considerations and experimental data suggest that a major mechanism of clearance of compounds placed into the peritoneal cavity is by way of the portal circulation. The delivery of the drug to the liver by way of the portal circulation essentially equals the amount of drug entering the liver during intrahepatic artery infusion.

Several clinical trials studied whether perioperative adjuvant portal vein injection in patients with colon cancer can prevent recurrences in the liver. In Taylor’s study, benefit was seen in patients with Dukes’ B. The Weber and Wolmark studies showed an increase of survival in patients who received adjuvant intraperitoneal infusion of chemotherapy after resection of their primary colon tumor. However, none of these studies demonstrated a decrease in the incidence of liver metastases in contrast to IHC, which demonstrates a significant decrease.

The reason for accelerated tumor growth after hepatectomy seems to be due to growth factors, such as hepatocyte growth factor and transforming growth factor-α, secreted from the residual liver of both humans and mice during the first week after a partial hepatectomy. In a previous study, we demonstrated an increase of human colon cancer growth implanted into the liver of nude mice after 30% hepatectomy. The timing of initiation of chemotherapy in our study is important for 2 reasons: 1. the potential effect of growth factors that can stim-

TABLE I - TREATMENT REGIMENS OF HUMAN HEPATOCELLULAR CARCINOMA IN THE LIVER OF NUDEN MICE

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of animals</th>
<th>Therapeutic regimens listed in order of use</th>
</tr>
</thead>
<tbody>
<tr>
<td>(1) Control</td>
<td>9</td>
<td>saline 0.5 ml, once a day for 5 days.</td>
</tr>
<tr>
<td>(2) 5-FU alone</td>
<td>8</td>
<td>20 mg/kg/0.5 ml once a day for 5 days; first dose 3 days after tumor implantation.</td>
</tr>
<tr>
<td>(3) Liver resection alone</td>
<td>10</td>
<td>curative liver resection 3 days after tumor implantation.</td>
</tr>
<tr>
<td>(4) Adjuvant</td>
<td>8</td>
<td>curative liver resection + 2 days rest + 5-FU 20 mg/kg/0.5 ml once a day for 5 days.</td>
</tr>
<tr>
<td>(5) Neoadjuvant</td>
<td>8</td>
<td>5-FU 20 mg/kg/0.5 ml once a day for 5 days + 2 days rest + curative liver resection.</td>
</tr>
<tr>
<td>(6) Intra-hepatectomy chemotherapy</td>
<td>10</td>
<td>5-FU 20 mg/kg/0.5 ml, first dose 2 hr prior to curative liver resection + 4 more doses beginning from post-operative first day.</td>
</tr>
</tbody>
</table>
ulate tumor growth released early after liver resection; 2. surgical manipulation spreads malignant cells into the portal circulation.\textsuperscript{12,19,29} The highest concentration of 5-FU after i.p. injection on liver tumors is reached after 2 hr.\textsuperscript{40} Therefore, in our study, in the IHC group, the first dose of 5-FU was injected i.p. 2 hr before the start of the operation. The other 4 doses were administered daily from the first post-operative day through day-4.

The IHC strategy significantly prolonged survival and in 5 of 10 animals prevented tumor recurrence. Despite the recurrences of tumor in the liver in 5 of the animals treated with IHC 5-FU, lymph node metastases draining the liver occurred in only 2 animals and peritoneal carcinomatosis in only 1 animal (Table III). The IHC thus seemingly reduced the malignancy of the tumor.

The data in our study demonstrated that IHC for resectable HCC is an effective and convenient procedure. IHC therapy does not have the disadvantage of neoadjuvant chemotherapy in which a resectable tumor remains in the liver parenchyma with the possibility of becoming nonoperable in case there is no response to chemotherapy.\textsuperscript{41} The results of this study indicate that IHC for resection of HCC should be investigated clinically.

TABLE II – SURVIVAL EFFICACY OF INTRA-HEPATECTOMY CHEMOTHERAPY COMPARED WITH STANDARD TREATMENT MODALITIES IN NUDE MICE WITH HCC

<table>
<thead>
<tr>
<th>Group</th>
<th>At the time when all mice in the control had died</th>
<th>At the time when all mice in the 5-FU group had died</th>
<th>At the time when all mice in the surgery group had died</th>
<th>At the time when all mice in the adjuvant group had died</th>
<th>At the time when all mice in the neoadjuvant group had died</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Survival rate (%)</td>
<td>p-value* vs. control</td>
<td>Survival rate (%)</td>
<td>p-value* vs. 5-FU</td>
<td>Survival rate (%)</td>
</tr>
<tr>
<td>Control</td>
<td>0</td>
<td>—</td>
<td>0</td>
<td>—</td>
<td>0</td>
</tr>
<tr>
<td>5-FU</td>
<td>20</td>
<td>0.474</td>
<td>0</td>
<td>—</td>
<td>0</td>
</tr>
<tr>
<td>Surgery</td>
<td>60</td>
<td>0.011</td>
<td>30</td>
<td>0.211</td>
<td>20</td>
</tr>
<tr>
<td>Adjuvant</td>
<td>30</td>
<td>0.211</td>
<td>20</td>
<td>0.474</td>
<td>20</td>
</tr>
<tr>
<td>Neoadjuvant</td>
<td>70</td>
<td>0.003</td>
<td>60</td>
<td>0.011</td>
<td>40</td>
</tr>
<tr>
<td>Intra-hepatectomy</td>
<td>80</td>
<td>0.001</td>
<td>70</td>
<td>0.003</td>
<td>70</td>
</tr>
</tbody>
</table>

*All p-values were derived from the $\chi^2$/Fisher exact test.
TABLE III – EFFICACY OF INTRA-HEPATECTOMY CHEMOTHERAPY COMPARED WITH STANDARD TREATMENT MODALITIES ON GROWTH AND METASTASIS OF HCC IN NUDE MICE

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of mice analyzed in each group</th>
<th>Mean tumor weight (g) + SD</th>
<th>p-value(^1) compared with the control</th>
<th>Lymph node metastasis</th>
<th>p-value(^2) compared with the control</th>
<th>Peritoneal carcinomatosis</th>
<th>p-value(^3) compared with the control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>9</td>
<td>8.8 ± 3.14</td>
<td>—</td>
<td>9/9</td>
<td>6/9</td>
<td>—</td>
<td></td>
</tr>
<tr>
<td>Surgery</td>
<td>8</td>
<td>6.9 ± 4.64</td>
<td>0.834</td>
<td>6/8</td>
<td>0.206</td>
<td>2/8</td>
<td>0.153</td>
</tr>
<tr>
<td>5-FU</td>
<td>10</td>
<td>9.1 ± 1.75</td>
<td>1.000</td>
<td>10/10</td>
<td>1.000</td>
<td>6/10</td>
<td>1.000</td>
</tr>
<tr>
<td>Adjuvant</td>
<td>8</td>
<td>6.8 ± 4.02</td>
<td>0.878</td>
<td>5/8</td>
<td>0.082</td>
<td>2/8</td>
<td>0.153</td>
</tr>
<tr>
<td>Neoadjuvant</td>
<td>8</td>
<td>7.1 ± 3.56</td>
<td>0.846</td>
<td>5/8</td>
<td>0.082</td>
<td>3/8</td>
<td>0.347</td>
</tr>
<tr>
<td>Intra-hepatectomy chemotherapy</td>
<td>10</td>
<td>2.7 ± 2.90</td>
<td>0.003</td>
<td>2/10</td>
<td>0.001</td>
<td>1/10</td>
<td>0.020</td>
</tr>
</tbody>
</table>

\(^1\)By the ANOVA test. \(^2\)By the \(\chi^2\)Fisher exact test.

REFERENCES

26. Taylor I, Machin D, Mullee M, Trotter G, Cooke T, West C. A


